

# TruSeq Rapid Exome Library Prep Protocol Guide

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|                                     |    |
|-------------------------------------|----|
| Tagment Genomic DNA                 | 3  |
| Clean Up Tagmented DNA              | 4  |
| Amplify Tagmented DNA               | 5  |
| Clean Up Amplified DNA              | 6  |
| Hybridize Probes                    | 7  |
| Capture Hybridized Probes           | 9  |
| Perform Second Hybridization        | 11 |
| Perform Second Capture              | 13 |
| Clean Up Captured Library           | 15 |
| Amplify Enriched Library            | 16 |
| Clean Up Amplified Enriched Library | 17 |
| Validate Enriched Libraries         | 18 |
| Acronyms                            | 19 |
| Technical Assistance                | 21 |



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# Tagment Genomic DNA

## Preparation

- 1 Save the following TAG58 program on the thermal cycler:
  - ▶ Choose the preheat lid option and set to 100°C
  - ▶ 58°C for 10 minutes
  - ▶ Hold at 10°C
  - ▶ Each well or tube contains 50  $\mu$ l.
- 2 Save the following TAG60 program on the thermal cycler:
  - ▶ Choose the preheat lid option and set to 100°C
  - ▶ 60°C for 5 minutes
  - ▶ Hold at 10°C
  - ▶ Each well or tube contains 65  $\mu$ l.

## Procedure

- 1 Quantify gDNA using a fluorometric method.
- 2 Dilute gDNA in Tris-HCl 10 mM, pH 8.5 to a final volume of 10  $\mu$ l at 5 ng/ $\mu$ l.
- 3 Add the following items in the order listed to a new Hard-Shell PCR plate or to a new 0.2 ml thin-wall PCR tube or 8-tube strip.
  - ▶ TD (25  $\mu$ l)
  - ▶ Normalized gDNA (10  $\mu$ l)
  - ▶ TDE2 (15  $\mu$ l)
- 4 Mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 5 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at 280  $\times$  g for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 6 Place on the preprogrammed thermal cycler and run the TAG58 program.
- 7 Add 15  $\mu$ l ST2, and then pipette to mix
- 8 Place on the preprogrammed thermal cycler and run the TAG60 program.

# Clean Up Tagmented DNA

## Procedure

- 1 Transfer all supernatant to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 2 Add 52  $\mu$ l SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Pipette up and down 10 times.
  - ▶ [Tube] Pipette up and down.
- 3 Incubate at room temperature for 5 minutes.
- 4 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 5 Transfer 98  $\mu$ l supernatant to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 6 Add 137  $\mu$ l SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Pipette up and down 10 times.
  - ▶ [Tube] Pipette up and down.
- 7 Incubate at room temperature for 5 minutes.
- 8 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 9 Remove and discard all supernatant.
- 10 Wash 2 times with 200  $\mu$ l 80% EtOH.
- 11 Using a 20  $\mu$ l pipette, remove residual 80% EtOH.
- 12 Air-dry on the magnetic stand for 5 minutes.
- 13 Add 22.5  $\mu$ l RSB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 14 Remove from the magnetic stand.
- 15 Incubate at room temperature for 2 minutes.
- 16 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 17 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 18 Transfer 20  $\mu$ l supernatant to a new Hard-Shell PCR plate or to a new 8-tube strip.

# Amplify Tagmented DNA

## Preparation

- 1 Save the following LAM AMP program on the thermal cycler:
  - ▶ Choose the preheat lid option and set to 100°C
  - ▶ 72°C for 3 minutes
  - ▶ 98°C for 30 seconds
  - ▶ 10 cycles of:
    - ▶ 98°C for 10 seconds
    - ▶ 60°C for 30 seconds
    - ▶ 72°C for 30 seconds
  - ▶ 72°C for 5 minutes
  - ▶ Hold at 10°C
  - ▶ Each well or tube contains 50  $\mu$ l.

## Procedure

- 1 [Plate] Arrange Index 1 (i7) adapters in columns 1–12 of the TruSeq Index Plate Fixture.
- 2 [Plate] Arrange Index 2 (i5) adapters in rows A–H of the TruSeq Index Plate Fixture.
- 3 [Plate] Place the plate on the TruSeq Index Plate Fixture.
- 4 Add 5  $\mu$ l of each Index 1 (i7) adapter as follows.
  - ▶ [Plate] Using a multichannel pipette, add to each column.
  - ▶ [Tube] Add a different index to each tube.
- 5 Replace the cap on each i7 adapter tube with a new orange cap.
- 6 Add 5  $\mu$ l of each Index 2 (i5) adapter as follows.
  - ▶ [Plate] Using a multichannel pipette, add to each column.
  - ▶ [Tube] Add a different index to each tube.
- 7 Replace the cap on each i5 adapter tube with a new white cap.
- 8 Add 20  $\mu$ l LAM, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 9 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 10 Place on the thermal cycler and run the LAM AMP program.

### SAFE STOPPING POINT

If you are stopping, seal the plate or cap the tube and store at 2°C to 8°C for up to 2 days. Alternatively, leave on the thermal cycler overnight.

# Clean Up Amplified DNA

## Procedure

- 1 Centrifuge at  $280 \times g$  for 1 minute.
- 2 Transfer 50  $\mu\text{l}$  total volume to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 3 Add 90  $\mu\text{l}$  SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 4 Incubate at room temperature for 5 minutes.
- 5 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 6 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 7 Remove and discard all supernatant.
- 8 Wash 2 times with 200  $\mu\text{l}$  80% EtOH.
- 9 Using a 20  $\mu\text{l}$  pipette, remove residual 80% EtOH.
- 10 Air-dry on the magnetic stand for 5 minutes.
- 11 Add 17  $\mu\text{l}$  RSB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 12 Remove from the magnetic stand.
- 13 Incubate at room temperature for 2 minutes.
- 14 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 15 Place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 16 Transfer 15  $\mu\text{l}$  supernatant to a new Hard-Shell PCR plate or to a new 1.5 ml microcentrifuge tube or 8-tube strip.
- 17 Quantify the library using a fluorometric method.

## SAFE STOPPING POINT

If you are stopping, seal the plate or cap the tube and store at  $-25^{\circ}\text{C}$  to  $-15^{\circ}\text{C}$  for up to 14 days.

# Hybridize Probes

## Preparation

- 1 Save the TRE HYB program on the thermal cycler:
  - ▶ Choose the preheat lid option and set to 100°C
  - ▶ 95°C for 10 minutes
  - ▶ 58°C for 30 minutes
  - ▶ Each well or tube contains 10  $\mu$ l.

## Pool Libraries

- 1 Combine 500 ng of each DNA library, making sure that each library has a unique index.
  - ▶ If the total volume is > 30  $\mu$ l, concentrate the pooled sample to 30  $\mu$ l.
  - ▶ If the total volume is < 30  $\mu$ l, increase the volume to 30  $\mu$ l with RSB.

## Procedure

- 1 Add the following reagents in the order listed to a new midi plate or to a new 1.5 ml microcentrifuge tube.
  - ▶ DNA library sample or pool (30  $\mu$ l)
  - ▶ BLR (10  $\mu$ l)
  - ▶ CEX (10  $\mu$ l)
- 2 Mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 3 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 4 Add 125  $\mu$ l SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 5 Incubate at room temperature for 10 minutes.
- 6 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 7 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 8 Remove and discard all supernatant.
- 9 Wash 2 times with 200  $\mu$ l 80% EtOH.
- 10 Using a 20  $\mu$ l pipette, remove residual 80% EtOH.
- 11 Air-dry on the magnetic stand for 10 minutes.

- 12 Add 7.7  $\mu$ l EHB1, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 13 Remove from the magnetic stand.
- 14 Incubate at room temperature for 2 minutes.
- 15 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 16 Place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 17 Transfer 7.5  $\mu$ l supernatant to a new Hard-Shell PCR plate or to a new thin-wall PCR tube or 8-tube strip.
- 18 Add 2.5  $\mu$ l EHB2, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 19 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 20 Place on the thermal cycler and run the TRE HYB program.



# Capture Hybridized Probes

## Preparation

- 1 [Plate] Preheat a microheating system with midi plate insert to 50°C.
- 2 [Tube] Preheat a heat block to 50°C.

## Procedure

- 1 Centrifuge at  $280 \times g$  for 1 minute.
- 2 Transfer all (~10  $\mu$ l) to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 3 Add 250  $\mu$ l SMB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 5 minutes.
  - ▶ [Tube] Pipette up and down.
- 4 Incubate at room temperature for 25 minutes.
- 5 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 6 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 7 Remove and discard all supernatant.
- 8 Remove from the magnetic stand.
- 9 Add 200  $\mu$ l EEW, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 4 minutes. Pipette to resuspend the bead pellet further.
  - ▶ [Tube] Pipette up and down.
- 10 Incubate as follows.
  - ▶ [Plate] Place on the 50°C microheating system with the lid closed for 30 minutes.
  - ▶ [Tube] Place on the 50°C heat block for 30 minutes.
- 11 Immediately place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 12 Remove and discard all supernatant.
- 13 Remove from the magnetic stand.
- 14 Repeat steps 9–13 for a total of 2 washes.
- 15 Mix 28.5  $\mu$ l EE1 and 1.5  $\mu$ l HP3, and then vortex.
- 16 Add 23  $\mu$ l elution premix, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 2 minutes.
  - ▶ [Tube] Pipette up and down.
- 17 Incubate at room temperature for 2 minutes.
- 18 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.

- 19 Place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 20 Transfer 21  $\mu$ l supernatant to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 21 Add 4  $\mu$ l ET2, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 22 Add 5  $\mu$ l RSB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 23 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.

### **SAFE STOPPING POINT**

If you are stopping, seal the plate or cap the tube and store at  $-25^{\circ}\text{C}$  to  $-15^{\circ}\text{C}$  for up to 7 days.

## Perform Second Hybridization

### Procedure

- 1 Add the following reagents in the order listed.
  - ▶ BLR (10  $\mu$ l)
  - ▶ CEX (10  $\mu$ l)
- 2 Mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 3 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 4 Add 125  $\mu$ l SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 5 Incubate at room temperature for 10 minutes.
- 6 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 7 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 8 Remove and discard all supernatant.
- 9 Wash 2 times with 200  $\mu$ l 80% EtOH.
- 10 Using a 20  $\mu$ l pipette, remove residual 80% EtOH.
- 11 Air-dry on the magnetic stand for 10 minutes.
- 12 Add 7.7  $\mu$ l EHB1, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 13 Remove from the magnetic stand.
- 14 Incubate at room temperature for 2 minutes.
- 15 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 16 Place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 17 Transfer 7.5  $\mu$ l supernatant to a new Hard-Shell PCR plate or to a new 8-tube strip.
- 18 Add 2.5  $\mu$ l EHB2, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 19 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.

20 Place on the thermal cycler and run the TRE HYB program.

## Perform Second Capture

### Preparation

- 1 [Plate] Preheat a microheating system with midi plate insert to 50°C.
- 2 [Tube] Preheat a heat block to 50°C.

### Procedure

- 1 Centrifuge at  $280 \times g$  for 1 minute.
- 2 Transfer 10  $\mu\text{l}$  supernatant to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 3 Add 250  $\mu\text{l}$  SMB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 5 minutes.
  - ▶ [Tube] Pipette up and down.
- 4 Incubate at room temperature for 25 minutes.
- 5 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 6 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 7 Remove and discard all supernatant.
- 8 Remove from the magnetic stand.
- 9 Add 200  $\mu\text{l}$  EEW, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 4 minutes. Pipette to resuspend the bead pellet further.
  - ▶ [Tube] Pipette up and down.
- 10 Incubate as follows.
  - ▶ [Plate] Place on the 50°C microheating system with the lid closed for 30 minutes.
  - ▶ [Tube] Place on the 50°C heat block for 30 minutes.
- 11 Immediately place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 12 Remove and discard all supernatant.
- 13 Remove from the magnetic stand.
- 14 Repeat steps 9–13 for a total of 2 washes.
- 15 Mix 28.5  $\mu\text{l}$  EE1 and 1.5  $\mu\text{l}$  HP3, and then vortex.
- 16 Add 23  $\mu\text{l}$  elution premix, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 2 minutes.
  - ▶ [Tube] Pipette up and down.
- 17 Incubate at room temperature for 2 minutes.

- 18 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 19 Place on a magnetic stand and wait until the liquid is clear (~2 minutes).
- 20 Transfer 21  $\mu\text{l}$  supernatant to a new midi plate or to a new 1.5 ml microcentrifuge tube or 8-tube strip.
- 21 Add 4  $\mu\text{l}$  ET2, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 22 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.

## Clean Up Captured Library

### Procedure

- 1 Add 45  $\mu$ l SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 2 Incubate at room temperature for 5 minutes.
- 3 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 4 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 5 Remove and discard all supernatant.
- 6 Wash 2 times with 200  $\mu$ l 80% EtOH.
- 7 Use a 20  $\mu$ l pipette to remove residual EtOH.
- 8 Air-dry on the magnetic stand until dry (~5 minutes).
- 9 Add 27.5  $\mu$ l RSB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 10 Remove from the magnetic stand.
- 11 Incubate at room temperature for 2 minutes.
- 12 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 13 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 14 Transfer 25  $\mu$ l supernatant to a new Hard-Shell PCR plate or to a new 8-tube strip.

### SAFE STOPPING POINT

If you are stopping, seal the plate or cap the tube and store at  $-25^{\circ}\text{C}$  to  $-15^{\circ}\text{C}$  for up to 7 days.

# Amplify Enriched Library

## Preparation

- 1 Save the following AMP10 program on the thermal cycler:
  - ▶ Choose the preheat lid option and set to 100°C
  - ▶ 98°C for 30 seconds
  - ▶ 10 cycles of:
    - ▶ 98°C for 10 seconds
    - ▶ 60°C for 30 seconds
    - ▶ 72°C for 30 seconds
  - ▶ 72°C for 5 minutes
  - ▶ Hold at 10°C
  - ▶ Each well or tube contains 50  $\mu$ l.

## Procedure

- 1 Add 5  $\mu$ l PPC.
- 2 Add 20  $\mu$ l EAM, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1200 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 3 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 4 Place on the thermal cycler and run the AMP10 program.

### SAFE STOPPING POINT

If you are stopping, seal the plate or cap the tube and store at 2°C to 8°C for up to 2 days. Alternatively, leave on the thermal cycler overnight.



## Clean Up Amplified Enriched Library

### Procedure

- 1 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 2 Transfer 50  $\mu\text{l}$  to a new midi plate or to a new 1.5 ml microcentrifuge tube.
- 3 Add 50  $\mu\text{l}$  SPB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 4 Incubate at room temperature for 5 minutes.
- 5 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 6 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 7 Remove and discard all supernatant.
- 8 Wash 2 times with 200  $\mu\text{l}$  80% EtOH.
- 9 Use a 20  $\mu\text{l}$  pipette to remove residual EtOH.
- 10 Air-dry on the magnetic stand until dry (~5 minutes).
- 11 Add 32  $\mu\text{l}$  RSB, and then mix thoroughly as follows.
  - ▶ [Plate] Shake at 1800 rpm for 1 minute.
  - ▶ [Tube] Pipette up and down.
- 12 Remove from the magnetic stand.
- 13 Incubate at room temperature for 2 minutes.
- 14 Centrifuge as follows.
  - ▶ [Plate] Centrifuge at  $280 \times g$  for 1 minute.
  - ▶ [Tube] Centrifuge briefly.
- 15 Place on a magnetic stand and wait until the liquid is clear (2–5 minutes).
- 16 Transfer 30  $\mu\text{l}$  supernatant to a new Hard-Shell PCR plate or to a new 1.5 ml microcentrifuge tube or 8-tube strip.

### SAFE STOPPING POINT

If you are stopping, seal the plate or cap the tube and store at  $-25^{\circ}\text{C}$  to  $-15^{\circ}\text{C}$  for up to 7 days.

# Validate Enriched Libraries

## Quantify Libraries

- 1 Quantify the postenriched library using the Qubit dsDNA BR Assay Kit.

## Assess Quality

- 1 If the library concentration is higher than the supported quantitative range for the High Sensitivity DNA chip, dilute the library 1:10 with RSB.
- 2 Run 1  $\mu$ l of post enriched library using a High Sensitivity DNA chip.

## Acronyms

| Acronym | Definition                        |
|---------|-----------------------------------|
| BLR     | Blocker                           |
| CEX     | Coding Exome Oligos               |
| EAM     | Enrichment Amplification Mix      |
| EE1     | Enrichment Elution Buffer 1       |
| EEW     | Enhanced Enrichment Wash Solution |
| EHB1    | Enrichment Hybridization Buffer 1 |
| EHB2    | Enrichment Hybridization Buffer 2 |
| ET2     | Elute Target Buffer 2             |
| HP3     | 2N NaOH                           |
| LAM     | Library Amplification Mix         |
| PPC     | PCR Primer Cocktail               |
| RSB     | Resuspension Buffer               |
| SMB     | Streptavidin Magnetic Beads       |
| SPB     | Sample Purification Beads         |
| ST2     | Stop Tagment Buffer 2             |
| TD      | Tagment DNA Buffer                |
| TDE2    | Tagment DNA Enzyme 2              |

## Notes

## Technical Assistance

For technical assistance, contact Illumina Technical Support.

**Table 1** Illumina General Contact Information

|                |                          |
|----------------|--------------------------|
| <b>Website</b> | www.illumina.com         |
| <b>Email</b>   | techsupport@illumina.com |

**Table 2** Illumina Customer Support Telephone Numbers

| Region        | Contact Number | Region          | Contact Number  |
|---------------|----------------|-----------------|-----------------|
| North America | 1.800.809.4566 | Japan           | 0800.111.5011   |
| Australia     | 1.800.775.688  | Netherlands     | 0800.0223859    |
| Austria       | 0800.296575    | New Zealand     | 0800.451.650    |
| Belgium       | 0800.81102     | Norway          | 800.16836       |
| China         | 400.635.9898   | Singapore       | 1.800.579.2745  |
| Denmark       | 80882346       | Spain           | 900.812168      |
| Finland       | 0800.918363    | Sweden          | 020790181       |
| France        | 0800.911850    | Switzerland     | 0800.563118     |
| Germany       | 0800.180.8994  | Taiwan          | 00806651752     |
| Hong Kong     | 800960230      | United Kingdom  | 0800.917.0041   |
| Ireland       | 1.800.812949   | Other countries | +44.1799.534000 |
| Italy         | 800.874909     |                 |                 |

**Safety data sheets (SDSs)**—Available on the Illumina website at [support.illumina.com/sds.html](http://support.illumina.com/sds.html).

**Product documentation**—Available for download in PDF from the Illumina website. Go to [support.illumina.com](http://support.illumina.com), select a product, then select **Documentation & Literature**.



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